Bioinformatics Lunch Seminar (Summer 2014)



- Every other Friday at 1 pm. 20-30 minutes plus discussion
- Informal, ask questions anytime, start discussions
- Content will be based on feedback
- Targeted at broad audience of various levels of backgrounds and education
- Emphasis on Genomics Center

Contact: Raymond Hovey Genomics Center - SFS <u>rhovey@uwm.edu</u> 414-382-1774 http://www.greatlakesgenomics.uwm.edu/



Pairwise sequence alignment

- Is the most fundamental operation of bioinformatics
- It is used to decide if two proteins (or genes) are related structurally or functionally
- It is used to identify domains or motifs that are shared between proteins
- It is used in the analysis of genomes

Overview of Sequence Alignment

Sequence alignment is the procedure of comparing two (pairwise alignment)

or

more (multiple sequence alignment) sequences by searching characters that are in the same <u>order</u> in the sequences. (order may not change, insertions and deletions are common, rearranging AAs is not)

Sequence alignment is useful for discovering functional, structural, and evolutionary information.

Sequences that are very much alike, or "similar" may have the same function.

Why Do We Align Sequences?

The basic idea of aligning sequences is that <u>similar sequences generally</u> produce similar proteins.

Therefore, if the sequences are similar they are *likely* to be functionally similar proteins.

To predict the characteristics of a protein using only its sequence data, sequence similarity of an unknown sequence with a protein of characterized function can be used to <u>identify the function of the unknown protein</u>

If one of them has known structure, then the alignment gives some <u>insight about</u> the structure of the other sequence.

How do we tell whether two macromolecules are similar?

Sequence Comparisons via Alignments provide an indication of similarity of two sequences.

Furthermore, sequence comparison and alignments permit to map functional information (such as the location of secondary structural elements, domains, active sites, and regulatory regions) present in a well-studied molecule to those that might be present in a new sequence. The **General Approach** involves the use of a set of algorithms such as **BLAST** programs to compare a query sequence to all the sequences in a specified database.

•Comparisons are made in a pairwise fashion.

•Each comparison is given a score reflecting the degree of similarity between the query and the sequence being compared.

•The higher the score, the greater the degree of similarity.

•The similarity is measured and shown by aligning two sequences.

•In general, alignments can be global or local (this is algorithm specific).

•A global alignment is an optimal alignment that includes all characters from each sequence, whereas a local alignment is an optimal alignment that includes only the **most similar local** region or regions.

•Discriminating between real and artifactual matches is done using an estimate of probability that the match might occur by chance.

•Similarity, by itself, cannot be considered a sufficient indicator of function but can be useful as a lead to understanding function.

Comparision that can be done (BLAST programs)

•DNA:DNA or RNA:RNA or DNA:RNA

polypeptide:polypeptide

•DNA translated in all 6 frames to polypeptide

•DNA translated in all 6 frames to all database DNA sequences translated in all 6 frames

Local vs. global alignments

Global Alignment

 Local Alignment—better alignment to find conserved segment

tccCAGTTATGTCAGgggacacgagcatgcagagac

aattgccgccgtcgttttcagCAGTTATGTCAGatc

Local vs. global alignments

Local Alignments

• Two genes in different species may be similar over short conserved regions and dissimilar over remaining regions.

Example:

Homeobox genes have a short region called the *homeodomain* that is highly conserved between species.

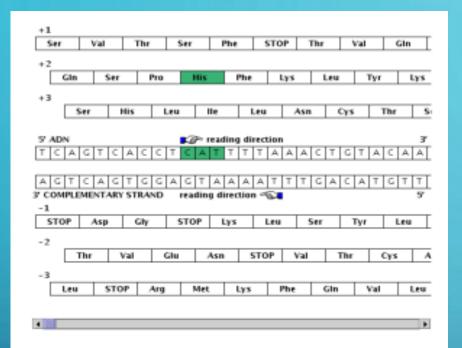
• A global alignment would not find the homeodomain because it would try to align the ENTIRE sequence.

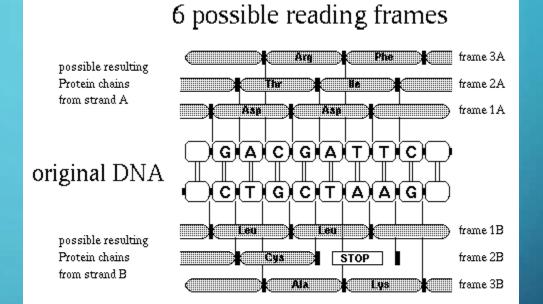
• Better for comparing sequences of different lengths. (like checking PCR primers against a genome)

Global alignment

- Finds best possible alignment across entire length of 2 sequences
- Aligned sequences assumed to be generally similar over entire length

DNA translated in all 6 frames





Pairwise Alignment

Global

- Best score from among alignments of full-length sequences
- Needleman-Wunsch algorithm (best alignment, dynamic programming)

Local

- Best score from among alignments of partial sequences
- Smith-Waterman algorithm (best alignment, dynamic programming)

http://www.ebi.ac.uk/Tools/psa/



The pros and cons of dynamic programming

These Algorithms will always give you the best possible alignment.

Due to their dynamic programming nature they are really slow and should not be used with multiple sequences (only pairwise).

Example:

It takes 100² seconds to perfectly align 2 protein Sequences of 100 aa each. Than it takes 100³ seconds to align 3 sequences. 100⁴ for 4 sequences 1.90258x10³⁴ Years for 20 sequences

BLAST (Biological Local Alignment Search Tool)

- BLAST programs use a heuristic search algorithm.
- The programs use the statistical methods of Karlin and Altschul.
- BLAST programs were designed for fast database searching, with minimal sacrifice of sensitivity for distantly related sequences.
- The programs search databases in a special compressed format.
- It is possible to use one's private database with BLAST. (required to convert it to the BLAST format)
- Great for comparing sequences of different lengths. (like checking PCR primers against a genome)

BLAST searching is fundamental to understanding the relatedness of any favorite query sequence to other known proteins or DNA sequences.

Why use **BLAST**?

Applications include

- identifying orthologs and paralogs
- discovering new genes or proteins
- discovering variants of genes or proteins
- investigating expressed sequence tags (ESTs)
- exploring protein structure and function

Orthologs are genes in different species that evolved from a common ancestral gene by speciation. Normally, orthologs retain the same function in the course of evolution.

Paralogs are genes related by duplication within a genome. Orthologs retain the same function in the course of evolution, whereas paralogs evolve new functions.

Four components to a BLAST search

(1) Choose the sequence (query)
(2) Select the BLAST program
(3) Choose the database to search
(4) Choose optional parameters

BLAST finds regions of similarity between biological sequences. more...

Learn more about how to use the new BLAST design

BLAST Assembled Genomes

Choose a species genome to search, or list all genomic BLAST databases

- Human
- Mouse
- <u>Rat</u>
- Arabidopsis thaliana

- Oryza sativa
 Bos taurus
- Danio rerio
- Drosophila melanogaster
- <u>Gallus gallus</u>
 Pan troglodytes
- Microbes
- Apis mellifera

Basic BLAST

Choose a BLAST program to run.

nucleotide blast	Search a nucleotide database using a nucleotide query						
Interestine must	Algorithms: blastn, megablast, discontiguous megablast						

protein blast Search protein database using a protein query Algorithms: blastp, psi-blast, phi-blast

blastx Search protein database using a translated nucleotide query

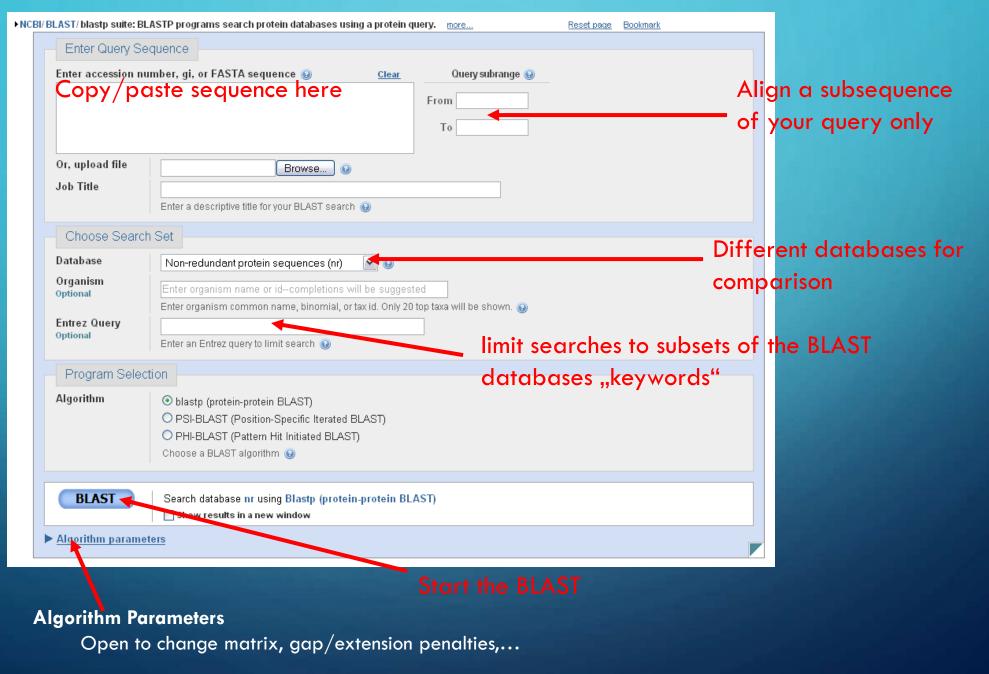
- tblastn | Search translated nucleotide database using a protein query
- tblastx | Search translated nucleotide database using a translated nucleotide query

Specialized BLAST

http://www.ncbi.nlm.nih.gov/BLAST/

To be discussed first

Protein-Protein BLAST (BLASTp): Structure of the NCBI web interface



Databases for BLAST (Example)

Non-redundant protein sequences (nr) Non-redundant protein sequences (nr) Reference proteins (refseq_protein) Swissprot protein sequences(swissprot) Patented protein sequences(pat) Protein Data Bank proteins(pdb) Environmental samples(env_nr)

Peptide Sequence Databases

nr

All non-redundant GenBank translations + RefSeq Proteins + PDB + SwissProt + PIR + PRF

0

sug

Only

refseq

non-redundant set of sequences at NCBI including genomic DNA, transcript

(RNA), and protein products, for major research organisms

swissprot

Last major release of the SWISS-PROT protein sequence database

pat

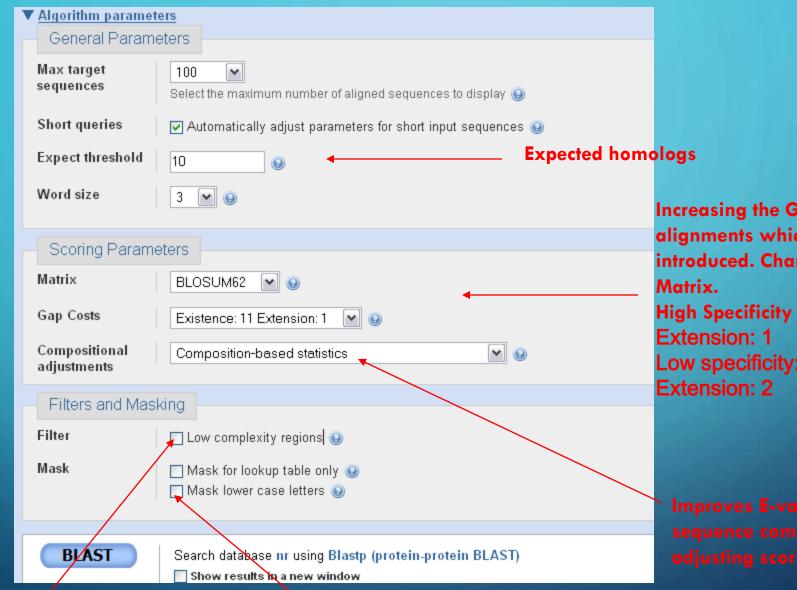
Proteins from the Patent division of GenPept.

pdb

Sequences derived from the 3-dimensional structure from Brookhaven Protein Data Bank

env_nr

Non-redudant database of environmental sequences



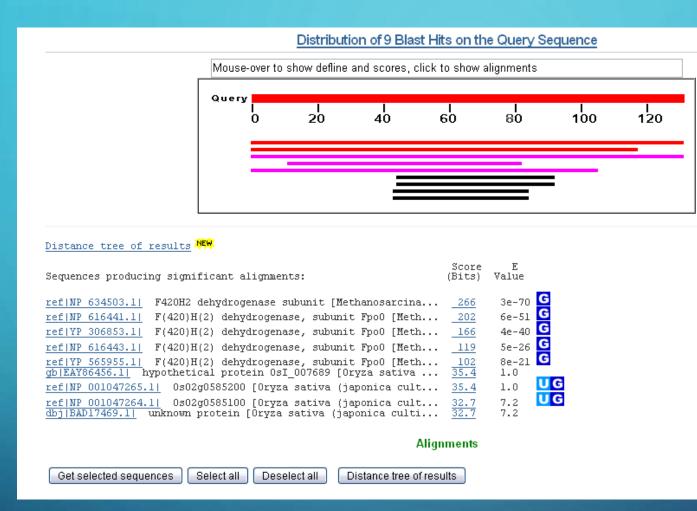
Increasing the Gap Costs will result in alignments which lower number of Gaps introduced. Changes automatically with Matrix. High Specificity PAM30 Existence: 9, Extension: 1 Low specificity: BLOSUM45 Existence: 15,

Improves E-value accuracy by taking query sequence complexity in consideration and adjusting scoring matrix.

Excludes sequences with low complexity

lenote areas you would like filtered with lower case

BLAST Output Example



Graphical Overview: An overview of the database sequences aligned to the query sequence is shown. The score of each alignment is indicated by one of five different colors, which divides the range of scores into five groups. At the bottom: Sequences with significant alignments

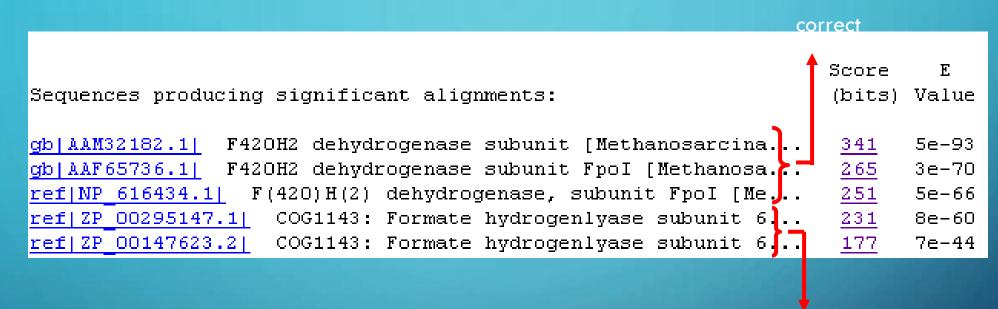


ref INP 634503.11 G F420H2 dehydrogenase subunit [Methanosarcina mazei Gol]
gb | AAF65742.1 | AF228525 12 G F420H2 dehydrogenase subunit Fpo0 [Methanosarcina mazei]
gb | AAM32175.11 G F420H2 dehydrogenase subunit [Methanosarcina mazei Gol]
Length=131

Score = 266 bits (679), Expect = 3e-70, Method: Composition-based stats. Identities = 131/131 (100%), Positives = 131/131 (100%), Gaps = 0/131 (0%)

Query	1	MTDCDLCGKGIPTVIPVRTYPPLLRFAYPEGVWKGLCETCLDSAQKTYLEVNRNHTSCRR MTDCDLCGKGIPTVIPVRTYPPLLRFAYPEGVWKGLCETCLDSAQKTYLEVNRNHTSCRR	60
Sbjct	l	MTDCDLCGKGIPTVIPVRTYPPLLRFAYPEGVWKGLCETCLDSAQKTYLEVNRNHTSCRR	60
Query	61	GKCSLCGSKTGVFSVELQIPDFSKGIVRKDVDVCYRCLKLVDEAYIRYKREQIEQDHEQG GKCSLCGSKTGVFSVELQIPDFSKGIVRKDVDVCYRCLKLVDEAYIRYKREQIEQDHEQG	120
Sbjct	61	GKCSLCGSKTGVFSVELQIPDFSKGIVRKDVDVCYRCLKLVDEAYIRYKREQIEQDHEQG	120
Query	121	RIHGHEHVHPH 131 RIHGHEHVHPH - Pairwise alignment	
Sbjet	121	RIHGHEHVHPH 131	

Pitfalls in Blast. Wrong annotation of genes/proteins!



not correct

Proteins are 98% identical First three annotations correct (from Methanosarcina mazei, Ms. acetivorans)– biochemical evidence No. 4 and 5 mis-annotated – corresponding protein from two closely related methanogenic archaea

Solution: use a curated database like SwissProt

Choose Searc	h Set
Database	Swissprot protein sequences(swissprot) 💽 💿
Organism Optional	Enter organism name or idcompletions will be suggested
Entrez Query Optional	Enter organism common name, binomial, or tax id. Only 20 top taxa will be shown. ⓐ Enter an Entrez query to limit search ⓐ

Statistical significance of Blast search: Matrices

The Matrix Option.

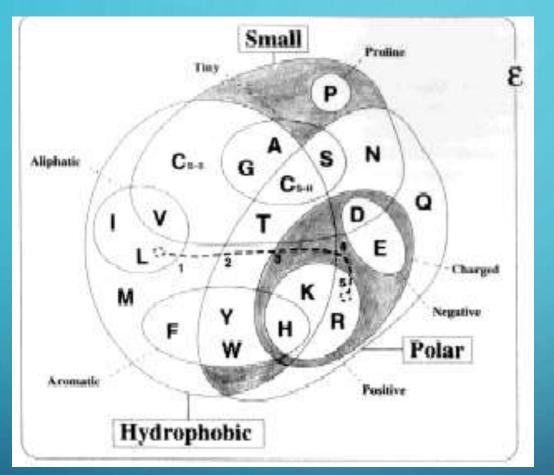
•A key element in evaluating the quality of a pairwise sequence alignment is the "substitution matrix", which assigns a score to aligning any possible pair of residues.

•In general, different substitution matrices are used to detect similarities among sequences that are diverged by differing degrees.

•A single matrix may nevertheless be reasonably efficient over a relatively broad range of evolutionary change.

•The strength of a match is determined by the returned score of the alignment.

How can we evaluate amino acid changes to score them adequately? -> look at properties of amino acids!



 $E \rightarrow D$ 1 border $E \rightarrow Q$ 2 borders

The 20 natural amino acids are characterized by a set of 10 physico-chemical properties (see above) and are organized according to the overlapping sectors. Lysin (K) for example is positively charged, polar and hydrophobic. Hence, there is a functional similarity to Arginine (R) and also a relationship to the polar and charged amino acid Glutamic Acid(E). To determine the degree of similarity in a position of the alignment the borders are counted that are crossed to connect the amino acid sequences.

Α	4																			
R	-1	5																		
Ν	-2	0	6		_															
D	-2	-2	1	6																
С	0	-3	-3	-3	9															
Q	-1	1	0	0	-3	5														
E	-1	0	0	2	-4	2	5													
G	0	-2	0	-1	-3	-2	-2	6												
Η	-2	0	1	-1	-3	0	0	-2	8											
Ι	-1	-3	-3	-3	-1	-3	-3	-4	-3	4										
L	-1	-2	-3	-4	-1	-2	-3	-4	-3	2	4									
K	-1	2	0	-1	-1	1	1	-2	-1	-3	-2	5								
Μ	-1	-2	-2	-3	-1	0	-2	-3	-2	1	2	-1	5							
F	-2	-3	-3	-3	-2	-3	-3	-3	-1	0	0	-3	0	6						
P	-1	-2	-2	-1	-3	-1	-1	-2	-2	-3	-3	-1	-2	-4	7					
S	1	-1	1	0	-1	0	0	0	-1	-2	-2	0	-1	-2	-1	4				
Τ	0	-1	0	-1	-1	-1	-1	-2	-2	-1	-1	-1	-1	-2	-1	1	5			
W	-3	-3	-4	-4	-2	-2	-3	-2	-2	-3	-2	-3	-1	1	-4	-3	-2	11		
Y	-2	-2	-2	-3	-2	-1	-2	-3	2	-1	-1	-2	-1	3	-3	-2	-2	2	7	
V	0	-3	-3	-3	-1	-2	-2	-3	-3	3	1	-2	1	-1	-2	-2	0	-3	-1	4
	Α	R	Ν	D	С	Q	E	G	Η	Ι	L	K	Μ	F	P	S	Τ	W	Y	V

ρ

 \bigcirc

Differences in score matrices

Par	Part of the Blosum62 matrix											
	A	R	Ν	D	С	Q	E	G	Н	I		
А	4	-1	-2	-2	0	-1	-1	0	-2	-1		
R	-1	5	0	-2	-3	1	0	-2	0	-3		
Ν	-2	0	6	1	-3	0	0	0	1	-3		
D	-2	-2	1	6	-3	0	2	-1	-1	-3		
С	0	-3	-3	-3	9	-3	-4	-3		-1		
Q	-1	1	0	0	-3	5	2	-2		-3		
E	-1	0	0	2	-4	2	5	-2	0	-3		
G	0	-2	0	-1	-3	-2	-2	6	-2	-4		
Н	-2	0	1	-1	-3	0	0	-2	8	-3		
İ	-1	-3	-3	-3	-1	-3	-3	-4	-3	4		

Part of the Blosum45 matrix

	A	R	Ν	D	С	Q	Ε	G	Н	I
A	5	-2	-1	-2	-1	-1	-1	0	-2	-1
R	-2	7	0	-1	-3	1	0	-2	0	-3
Ν	-1	0	6	2	-2	0	0	0	1	-2
D	-2	-1	2	7	-3	0	2	-1	0	4
С	-1	-3	-2	-3	12	ç	-3	-3	-3	-3
Q	-1	1	0	0	-3	6	2	-2	1	-2
E	-1	0	0	2	-3	2	6	-2	0	-3
G	0	-2	0	-1	-3	-2	-2	7	-2	-4
Н	-2	0	1	0	-3	1	0	-2	10	-3
Ι	-1	-3	-2	-4	-3	-2	-3	-4	-3	5

Different values for substitutions (and gaps)

Common Substitution Matrix Families

•PAM (Point Accepted Mutation)

Amino acid scoring matrices are traditionally PAM matrices. PAM40 is most sensitive for similar sequences. PAM250 is for more distantly related sequences.

BLOSSUM (Blocks Substitution Matrix)

•BLOSSUM matrices are most sensitive for local alignment of related sequences.

•BLOSUM62 is optimised for general BLASTP searches, and is suitable for most situations; it will recognise some amino acid substitutions as conservative (e.g. Arg to Lys). It is a good general matrix, set by <u>default for protein BLAST searches</u>

•Caution! You cannot compare the alignment scores (see later) from one matrix directly against the alignment scores from another matrix!

The following matrices are roughly equivalent...
PAM100 ==> Blosum90
PAM120 ==> Blosum80
PAM160 ==> Blosum60
PAM200 ==> Blosum52
PAM250 ==> Blosum45

Statistical significance of Blast search: Scores and E-values

Once BLAST has found a similar sequence to the query in the database, it is helpful to have some idea of whether the alignment is "good" (possible biological relationship), or whether the similarity observed is by chance alone. BLAST uses statistical theory to produce a bit score and expect value (E-value) for each alignment pair.

The raw score (S) gives an indication of how good the alignment is; the higher the score, the better the alignment. The score is calculated from a formula that takes into account the alignment of similar or identical residues, as well as any gaps introduced to align the sequences. Bit scores (S') are normalized, which means that the bit scores from different alignments can be compared, even if different scoring matrices have been used.

The E-value gives an indication of the statistical significance of a given pairwise alignment. The lower the Evalue, the more significant the hit. A sequence alignment that has an E-value of 0.05 means that this similarity has a 5 in 100 (1 in 20) chance of occurring by chance alone. *E* values below 10⁻⁶ are most probably statistically significant. *E* values above 10⁻⁶ but below 10⁻³ deserve a second look. *E* values above 10⁻³ should not be considered. Alignments:

Third hit gives score = 442 and e-value of 1e-122

> <mark>gi 68212169 ref ZP_00564014.1 </mark> conserved hypothetical protein [Methanococcoides burtonii DSM 6242]
<u>qi 68184324 qb EAM99061.1 </u> conserved hypothetical protein [Methanococcoides burtonii DSM 6242] Birren
Bit score Length=457 Raw score
Score = 442 bits (1137), Expect = 1e-122 Identities = 218/445 (48%), Positives = 296/445 (66%), Gaps = 11/445 (2%)
Query 5 AGAAEPSGPGDFTSNQFSKSGICSNCHGSSFGEWAGSMHSLADSDFFYNAMLQEYGVAAE 64 A A +P+G + S F + C+NCH + GSMH+ A SD Y +E +A+E
Sbjet 17 ASAVDPTGFDELNSEDFVTNSKCANCHAILRSQHEGSMHAFAYSDPLYQKEALLASE 73

De	ter	min	ati	on	of	sco	res	w	ith Blosum 62
А	G	А	А	Е	Р	S	G	Р	input sequence
А	S	А	V	D	Р	Т	G	F	homolog
4	0	4	0	2	7	1	6 -	-4	Sum = Score S

BLAST for Protein query sequences

blastp = Compares a protein sequence with a protein database

If you want to find something about the function of your protein, use blastp to compare your protein with other proteins contained in the databases; identify common regions between proteins, or collect related proteins (phylogenetic analysis;

tblastn = Compares a protein sequence with a nucleotide database

If you want to discover new genes encoding proteins (from multiple organisms), use tblastn to compare your protein with DNA sequences translated into their six possible reading frames; map a protein to genomic DNA;

BLAST for DNA query sequences

blastn = Compares a DNA sequence with a DNA database;

Mapping oligonucleotides, cDNAs, and PCR products to a genome; annotating genomic DNA; screening repetitive elements; cross-species sequence exploration;

tblastx = Compares a DNA translated into protein with a DNA database translated into protein;

Cross-species gene prediction at the genome or transcript level (ESTs); searching for genes not yet in protein databases;

blastx = Compares a DNA translated into protein with a protein sequence database;

Finding protein-coding genes in genomic cDNA; determining if a cDNA corresponds to a known protein;

• Use the latest database version.

• As a rule of thumb, run BLAST first, then depending on your results run a finer tool (BLAT, Ssearch, Smith Waterman, Blocks, etc.).

• Whenever possible, use protein or translated nucleotide sequences.

• Splitting large query sequences can help (> 1000 for DNA, > 200 for protein).

• If the query has repeated segments, remove them and repeat the search.



The UCSC and BLAT

- BLAT = <u>B</u>last <u>Like A</u>lignment <u>T</u>ool
- Optimized for shorter reads (primers and next generation sequences)
- Faster but less homology depth.
- Can link directly into the UCSC browser (and other hosted databases like The Cancer Genome Atlas)
- Supports large amount of queries

https://genome.ucsc.edu/FAQ/FAQblat.html

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Statistical significance of Blast search: Scores and E-values

Determined by statistical algorithms: **SCORE** S reflects the similarity of proteins/ genes and is based on the scoring matrix

S (raw score) = Sum of substitutions and gap scores (penalties)

S' (bit score) = normalized raw score that allows direct comparison of scores from different alignments, scoring matrices in different Blast searches.

S'=(lambdaS-lnK)/ln2

(λ and K given at the bottom of all blast searches)

E-value: expected value = number of alignments that are expected to occur by change = probability of false-positive results

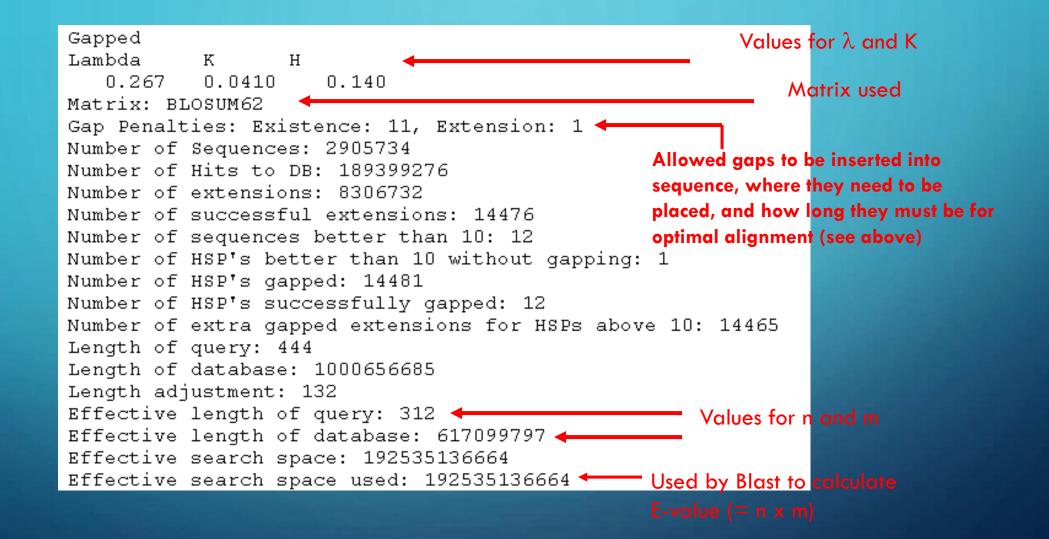
The E-value is an estimate of how many sequences would score by chance in the database searched. The higher E the higher the probablity that the aligment is made by chance. So we should look to low E values: Rule of thumb: E-values significant below 10^{-3} to 10^{-5}

The E- value is directly connected to S'.

 $E = mn 2^{-S'}$

(mn = effective search space; with m = effective length of query; n = effective length of database --- numbers given at the bottom of all blast searches).

P-Value: other value representing the significance of an alignment $P = 1 - e^{-E} - P < 0.05$ traditionally used to define statistical significance Statistical parameters of blast search (at the end of the results)



ρ